

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/27794932>

Superoxide dismutase phenotypes in duodenal ulcers: A genetic marker?

Article in *Indian Journal of Human Genetics* · September 2006

DOI: 10.4103/0971-6866.29855 · Source: OAI

CITATIONS

3

READS

80

5 authors, including:



Madhavi Jangala

MAA Research Foundation

18 PUBLICATIONS 113 CITATIONS

[SEE PROFILE](#)



Ananthapur Venkateshwari

Institute of Genetics and Hospital for Genetic Diseases, Osmania University, ...

131 PUBLICATIONS 728 CITATIONS

[SEE PROFILE](#)



Yasmeen Sheik

InterpretOmics India Pvt Ltd

6 PUBLICATIONS 16 CITATIONS

[SEE PROFILE](#)



Pratibha Nallari

Osmania University

208 PUBLICATIONS 1,606 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Genomics of Cardiomyopathy in Indian Populations [View project](#)



PH.D. WORK [View project](#)

Superoxide dismutase phenotypes in duodenal ulcers: A genetic marker?

S. Sulekha, J. Madhavi, A. Venkateshwari, S. Yasmeen, N. Pratibha
Department of Genetics, Osmania University, Hyderabad - 500 007, India

BACKGROUND: Cu-Zn superoxide dismutases are antioxidative defensive enzymes that catalyze the reduction of superoxide anions to hydrogen peroxide.

AIM: The study focuses on the association of electromorph of superoxide dismutase with duodenal ulcers, which result due to an imbalance between aggressive and defensive factors.

MATERIALS AND METHODS: Endoscopically confirmed 210 duodenal ulcer patients and 185 healthy individuals for comparative analysis were considered for the present study. Phenotyping of superoxide dismutase was carried out by subjecting the RBC membranes to polyacrylamide gel electrophoresis, using appropriate staining protocols.

RESULTS: Statistical analysis of SOD phenotypes revealed a significant increase of SOD A*2 allele and Superoxide dismutases (SOD) 2-2 phenotype in duodenal ulcer group. Among these individuals, a predominance of *Helicobacter pylori* infection was observed. The increased preponderance of homozygotes can be explained on the basis of reduced and altered enzyme activity, which may lead to disturbance in homeostasis of antioxidant/oxidant culminating in high lipid peroxidative gastric mucosal tissue damage and ulceration. No variation in the distribution of SOD phenotypes with respect to *Helicobacter pylori* indicates the role of Mn-SOD rather than Cu-Zn SOD in the *Helicobacter pylori* infected cases as reported earlier.

CONCLUSIONS: Superoxide dismutase as a genetic marker / gene modifier, encoding for an antioxidant enzyme in maintaining tissue homeostasis of the gastric mucosa is discussed.

Key words: Antioxidant, duodenal ulcer, free radical, gene modifier, genetic marker, lipid peroxidation, superoxide dismutase

of several aggressive factors like *Helicobacter pylori* infection, acid, pepsin and other proteases secreted on one hand and several defensive factors like bicarbonate and antiprotease on the other hand. Oxidative stress also plays an important role in vascular physiology and pathology.^[1] Previous studies have shown that superoxide anion is produced by oxidases present in human vascular cells.^[2] However, net levels of oxidative stress are determined in parallel by production and biodegradation of free radicals as well as by genetic susceptibility. The major anti-oxidant systems in the mucosal vasculature include, catalase, glutathione peroxidase, thioredoxin and superoxide dismutases, which modulate the oxidative stress but the primary role in the metabolism of superoxide anion radical is exerted by superoxide dismutases.^[2]

Superoxide dismutases (SOD) is well known for scavenging Superoxide radicals such as reactive oxygen species (ROS), subsequently protecting cells from oxidative injury and for maintaining tissue homeostasis. SOD is polymorphic in nature with three identifiable phenotypes SOD A*1, SOD A*2-1 and SOD A*2 encoded by two allelic forms SOD A*1 and SOD A*2 respectively and inherited as co-dominant alleles.^[3] The gene is localized to 21q21-q22.1 region.^[4] Since no information with regard to SOD and its electromorph association is available with respect to duodenal ulcers, such a study is imperative to identify the role of susceptible marker and its associated allele, apart from its role as a modifier gene in the genetic etiology of duodenal ulcers. Therefore, the present study aims at identifying specific electromorph association of SOD in duodenal ulcers with respect to *Helicobacter pylori* and its possible role in the etiopathogenesis of duodenal ulcers.

Introduction

Duodenal ulcers (DU) constituting the predominant form of peptic ulcers, found to occur in about 10% of the worldwide population, are characterized by the loss of mucosal resistance, as a result of action and interaction

Materials and Methods

Blood samples from endoscopically confirmed cases comprising of 210 duodenal ulcers and 185 healthy controls matched for age and sex, were collected from Department of Gastroenterology, Osmania General Hospital, Hyderabad and voluntary blood donor camps, Hyderabad.

Five ml of venous blood was collected in EDTA to obtain red blood cell membrane. The whole blood was washed with normal saline thrice and then centrifuged with ice-cold water to obtain red cell membranes for the phenotyping of superoxide dismutase. The *Helicobacter pylori* infection status was tested by rapid urease test in mucosal tissue biopsies of the duodenal ulcers as described by Vaira *et al.*^[5]

The phenotyping of SOD was based on Davies protocol^[6] of an 8% polyacrylamide gel electrophoresis gel and the electrophoresis was carried out using 0.005 M tris glycine running buffer at 4°C for two hours at 20 mA constant current. The gel was then placed in a tray and incubated in freshly prepared staining solution which consisted of 5 ml of phosphate buffer (pH 7.8), 8.5 ml of riboflavin, 10 mg of nitroblue tetrazolium chloride and 220 µl of TEMED prepared in 50 ml distilled water. Later, the gel was washed and transferred to distilled water and exposed under a bright fluorescent light till the background was blue and clear achromatic bands were visualized as cited by Beauchamp and Fridovich.^[7] Based on their mobility in the gel, the phenotypes of SOD A

were identified as 2-2, 2-1 and 1-1, respectively.

Frequency distribution was computed and allelic frequencies of SOD phenotypes were estimated to test for the Hardy Weinberg equilibrium^[8] and odds ratio (2 x 2 contingency) at 95% confidence interval limits was carried out to interpret the results statistically.^[9]

Results

Table 1 shows the frequency distribution of superoxide dismutase electromorphs in control and disease groups wherein SOD 2-1 phenotypes were found to be predominant in normal individuals (86%) while 2-2 phenotypic individuals are at an increased risk to the disease condition (94%).

The distribution of superoxide dismutase genotypic and allelic frequencies in control and duodenal ulcer groups is shown in Table 2. The frequency of SOD 2 and SOD 1 alleles were found to be 0.55 and 0.45 in healthy subjects and 0.96 and 0.04 in duodenal ulcer patients. A significant deviation from the Hardy-Weinberg equilibrium was observed in duodenal ulcer group strengthening the earlier observation [Table 1].

The odds ratio estimates of superoxide dismutase phenotypes in duodenal ulcer groups in comparison to control group, wherein an increased risk of SOD 2-2 phenotypic individuals to ulceration (log OR 5.6399) was observed, further strengthening the above observation [Table 3]. This could be attributed to SOD A*2 allele encoding for a less stable enzyme as a result of which altered scavenging of the superoxide free radicals may lead to oxidative stress induced mucosal damage, as seen in duodenal ulcers. Decreased activity of Cu-Zn SOD is also associated with initiation of nitric oxide (NO⁻) mediated apoptotic cell death and hence tissue damage in ulcers groups.^[10]

Table 4 shows the phenotypic distribution of SOD with respect to *Helicobacter pylori* status in control and

Table 1: Frequency distribution of superoxide dismutase electromorphs in control and duodenal ulcer groups

SOD A* phenotype	Control		Duodenal ulcers	
	nn	%		%
2-2	14	7.5	197	94.0
2-1	160	86.5	8	4.0
1-1	11	6.0	5	2.0
Total	185	100	210	100

Table 2: Genotypic and allelic frequency distribution of superoxide dismutase in control and duodenal ulcer groups

SOD A*Alleles	Allelic frequency		Types	Genotypic frequency	
	Control	Duodenal ulcer		Control	Duodenal ulcer
SOD 2A*	0.51	0.96	2-2	0.26	0.92
SOD1A*	0.49	0.04	2-1	0.24	0.0018
			1-1	0.50	0.082

Table 3: Odds test of association of SOD phenotypes in duodenal ulcers in comparison to control group

SOD phenotypic comparisons		Controls	Duodenal ulcers	Risk ratio	Odds ratio	Log odds	95% Confidence interval
2-2 vs 2-1	2-2	14	197	19.6066	281.4286*	5.6399*	115.1814 to 687.6287
	2-1	160	8				
2-2 vs 1-1	2-2	14	197	2.9877	30.9571*	3.4326*	9.436214 to 101.56
	1-1	11	5				
2-1 vs 1-1	2-1	160	8	0.1524	0.11	-2.2073	0.030784 to 0.393062
	1-1	11	5				
2-2 vs others	2-2	14	197	13.2147	185.0934*	5.2209*	84.62102 to 404.4912
	Others	171	13				
2-1 vs others	2-1	160	8	0.0535	0.0062	-5.0851	0.002723 to 0.014116
	Others	25	202				
1-1 vs others	1-1	22	5	0.5777	0.3858	-0.9524	0.131505 to 1.131837
	Others	174	205				

*Odds ratio >1 and lying between confidence interval with lower limit above 1 were taken as significant.

Table 4: Phenotypic distribution of superoxide dismutase with respect to *Helicobacter pylori* status in control and disease groups

SOD A*	Controls				Duodenal			
	HP+Ve		HP-Ve		HP+Ve		HP-Ve	
	n	%	n	%	n	%	n	%
2-2	10	32.2	1	1.3	175	93.0	22	100
2-1	20	64.4	72	91.0	8	4.3	-	-
1-1	1	3.2	6	7.6	5	2.6	-	-
Total	31	28.0	79	72.0	188	90.0	22	10.0

disease patients. In general, an interesting observation made is the predominance of SOD 2-2 phenotype in Hp +ve healthy subjects and ulcer patients, but the nature of association cannot be demonstrated statistically as the frequency distribution of 2-1 and 1-1 phenotypes in Hp-ve cases accounted to zero, further supporting the role of SOD as a gene modifier in such *Helicobacter pylori* infected cases. However, by observation, role of SOD A* 2 allele scavenging the O²⁻ derived free radical released due to mucosal damage as a consequence of *Helicobacter pylori* infection further justifies the SOD as a genetic marker as well as modifier of the disease in such Hp +ve cases. The study thus substantiates the role of SOD in the etiopathogenesis of duodenal ulcers and helps in genetic heterogeneity and risk prediction of duodenal ulcer groups.

Discussion

Cells have elaborated protective mechanisms to cope with potentially damaging molecules such as reactive oxygen species by synthesizing antioxidant enzymes.^[10] Superoxide dismutases (SOD) are key cellular defense

systems that disproportionate O²⁻ into oxygen and H₂O₂, with the latter being detoxified by glutathione peroxidase or catalase. Cu-Zn SOD referred to as SOD 1 and Mn SOD as SOD 2 are the two eukaryotic SOD's. Cu-Zn SOD located in the cytosol, is expressed constitutively and is considered as a housekeeping enzyme; in contrast to Mn SOD which is located in the mitochondrion. The Cu-Zn SOD enzyme is a known polymorphic marker encoded by two allelic forms SOD A*1 and SOD A*2 and inherited as co dominant alleles. An increased risk of SOD 2-2 phenotypic individuals to ulceration (OR: 281.43* CI: 115.18 to 687.68) was observed, this could be explained on the basis of SOD A*2 allele contributing to decreased stability of the enzyme or lowered enzyme synthesis. Studies by Novak *et al*^[11] revealed significant reduction in SOD activity and hence the locus might be 'regulatory' in nature wherein the altered structure and function of cell membranes especially of gastric mucosa as a result of ulceration, may have a key pathogenetic role in ulcers. SOD, a key enzyme in gastric mucosal protection, is depleted significantly in the ulcer edge compared to the controls and found to increase after healing with treatment.^[12] Hence, significant association of the SOD A*2 allele encoding for less stable enzyme, as a gene modifier in duodenal ulcers is justifiable.

Further, SOD A*2 allele contributing to less stable enzyme can have direct effect on lipid peroxidation and hence pathological progression of the disease. Significant elevation of malondialdehyde (MDA) in duodenal ulcers was reported which strengthens the

oxidative stress mechanism associated with duodenal ulcers.^[13] Since MDA levels are negatively correlated any altered function of SOD enzyme may in turn lead to increased MDA levels, disrupting the homeostatic balance causing high lipid peroxidation of the membranes of gastric mucosa^[14,15] which may culminate to ulceration, due to depletion of SOD levels.

Previous studies suggest that only Mn-SOD plays an important role as an antioxidant against ROS and, sub-sequently, in the maintenance of cell turnover in gastric mucosa.^[16] Gotz *et al* have shown that gastric tissue of *Helicobacter pylori* positive individuals was higher in activity and content of the cytokine inducible Mn-SOD, with no change in the constitutive Cu-Zn SOD.^[17] Hence Cu-Zn SOD role is rather trivial in case of *Helicobacter pylori* infection in duodenal ulcers. Our results are strengthened by the above observation implicating no association of *Helicobacter pylori* infection and Cu-Zn SOD enzyme.

Conclusion

The study highlights a significant risk of homozygous SOD A*2-2 phenotypic individuals to duodenal ulcers and SOD A*2 allele encoding for a less stable enzyme may lead to decreased scavenging of O²⁻ radicals resulting in the oxidative stress induced gastric mucosal tissue damage and ulcerogenesis. Thus SOD may be considered as a genetic marker in genetic risk prediction and may have a direct or modifier role in ulcerogenesis and could help in delineating the genetic heterogeneity of duodenal ulcers.

References

1. Tandon R, Khanna HD, Dorababu M, Goel RK. Oxidative stress and antioxidants status in peptic ulcer and gastric carcinoma. *Indian J Physiol Pharmacol* 2004;48:115-8.
2. Fridovich I. Superoxide dismutases. *Ann Rev Biochem* 1975;44:147-59.

3. DeCruo S, Kamboh MI, Leppert M, Ferrell RE. Isoelectric focusing of superoxide dismutase: Report of the unique SOD A*2 allele in a US white population. *Hum Hered* 1988;38:1-7.
4. Huret JL, Delabar JM, Marlhens F, Aurias A, Nicole A, Berthier M, *et al*. Down syndrome with duplication of a region of chromosome 21 containing the CuZn superoxide dismutase gene without detectable karyotypic abnormality. *Hum Genet* 1987;75:251-7.
5. Vaira D, Holtonb J, Menegattia M, Landia F, Riccia C, Ali A, *et al*. Blood tests in the management of *Helicobacter pylori* infection. *Gut* 1998;43:S39-46.
6. Davies BJ. Disc electrophoresis II. Method and application to human serum proteins. *Ann N Y Acad Sci* 1964;121:404-27.
7. Beauchamp C, Fridovich I. Superoxide dismutase: Improved assays and an assay applicable to acrylamide gels. *Anal Biochem* 1971;44:276-87.
8. Emery AE. *Statistical Methods in Medical Genetics*. Churchill Livingstone: UK; 1984.
9. Christensen R. *Loglinear models and logistic regression*. 2nd ed. Springer-Verlag: New York; 1951.
10. Brockhaus F, Brune B. Overexpression of CuZn superoxide dismutase protects RAW 264.7 macrophages against nitric oxide cytotoxicity. *Biochem J* 1999;338:295-303.
11. Novak R, Bosze Z, Matkovics B, Fachtet J. Gene affecting superoxide dismutase activity linked to the histocompatibility complex in H-2 congenic mice. *Science* 1980;207:86-7.
12. Klinowski E, Broide E, Varsano R, Eshchar J, Scapa E. Superoxide dismutase activity in duodenal ulcer patients. *Eur J Gastroenterol Hepatol* 1996;8:1151-5.
13. Venkateshwari A, Prathibha N, Vidyasagar A, Pratap B, Murty JS. Malondialdehyde levels in patients with duodenal ulcer. *Indian J Gastroenterol* 1995;14:57-8.
14. Wang SH, Wang YZ, Zhang KY, Shen JH, Zhou HQ, Qiu XY. Effect of superoxide dismutase and malondialdehyde metabolic changes on carcinogenesis of gastric carcinoma. *World J Gastroenterol* 2005;11:4305-10.
15. Zimmermann R, Flohe L, Weser U, Hartmann HJ. Inhibition of lipid peroxidation in isolated inner membrane of rat liver mitochondria by superoxide dismutase. *FEBS Lett* 1973;29:117-20.
16. Noguchi K, Kato K, Moriya T, Suzuki T, Saito M, Kikuchi T, *et al*. Analysis of cell damage in *Helicobacter pylori* associated gastritis. *Pathol Int* 2002;52:110-8.
17. Gotz JM, Thiol JL, Verspaget HW, Offeerhaus GJ, Biemond I, Lamers CB, *et al*. Treatment of *Helicobacter pylori* infection favourably affects gastric mucosal superoxide dismutase. *Gut* 1997;40:591-6.

Source of Support: Financial support from Indian Council of Medical Research, New Delhi is kindly acknowledged,
Conflict of Interest: None declared.